Next-generation whole-exome sequencing contribution to identification of rare autosomal recessive diseases

Tautvydas Rančelis,

Loreta Cimbalistienė,

Vaidutis Kučinskas

Department of Human and Medical Genetics, Faculty of Medicine, Vilnius University, Vilnius, Lithuania A rare disease is any disease that affects a small percentage of the population. In the European Union a disease is defined as rare if it affects less than 1 in 2,000 people. Despite a small percentage of affected people by one disease, the total number of rare diseases is estimated to be around 7,000–8,000, thus, because of their large number they have an impact on many people and even 30 million of European Union citizens may be suffering from them. Research of rare diseases may help to explain their mechanism or to develop more advanced diagnostics. Classical strategies for studies of rare autosomal recessive diseases encounter with additional problems (multiple genetic variants, de novo mutations, extremely rare cases) that make these strategies not enough effective. Next generation whole-exome sequencing (WES) opened a new page in Mendelian disease gene discovery – enabling to study autosomal recessive diseases in a new way. During 3 years of WES usage many novel mutations of autosomal recessive disease genes were discovered.

Key words: rare autosomal recessive diseases, whole exome sequencing, syndromes

INTRODUCTION

A rare disease is any disease that affects a small percentage of the population. In the European Union, a disease is defined as rare when it affects less than 1 in 2,000 people. The number of the rare diseases is estimated to be about 7,000–8,000 and 1,139 of them have been characterized as recessive. Approximately 80% have a defined genetic basis. Despite low frequency of diseases, because of their great number, even 30 million European Union citizens may be affected by rare diseases (1, 2). Considering this, research of these diseases has great importance and study of them may help to explain the mechanisms of diseases and help to develop more advanced diagnostics.

Classical strategies to identify the cause of Mendelian diseases rely on using linkage analysis, homozygosity mapping, and association analysis.

Correspondence to: Tautvydas Rančelis, Department of Human and Medical Genetics, Faculty of Medicine, Vilnius University, Santariškių 2, LT-08661 Vilnius, Lithuania. E-mail: tautvydas.rancelis@mf.vu.lt

Linkage analysis is family-based approach that was a great tool for Mendelian disease discovery and has a relatively long history of successful detection of the rare Mendelian diseases such as Huntington's disease and cystic fibrosis (3). However, some of the rare Mendelian diseases can be caused by multiple genetic variants as well as by new de novo mutations which, according to Vissers (4), can be linked to 7 of 10 patients of mental disability – in these cases linkage analysis is not effective.

Autozygosity mapping has been a good method for the identification of the autosomal recessive disease genes. However, that approach is limited by the unavailability of suitable consanguineous families' genealogies. While rare autosomal recessive diseases are overrepresented in consanguineous families, a significant proportion of affected patients nonetheless originate in families where the parents are apparently unrelated (5). So, even if there is a successful history of the rare disease identification with classical approaches, it encounters with problems like multiple genetic variants, de novo mutations, extremely rare cases that make these strategies not enough suitable for disease study. A new turning point was appearance of the next-generation sequencing (NGS) technologies.

NGS technologies are a powerful tool for identification of genetic disease causes. These technologies enable the whole genome and exome sequencing to avoid homozygosity mapping, prioritize candidate genes and to proceed directly to the examination of the list of variants. That gives enough insight to "pinpoint" the mutation responsible for a disease. At present several national projects using NGS are released in Canada, Korea, UK, USA, and other countries (6, 7).

Disease gene identification challenge using NGS technology changes to interpretation challenge, because using it researchers obtain a sizeable amount of variation data that must be thoroughly filtered. This part of NGS is the most difficult one and requires efficient tools for filtering and interpreting results of sequencing (8).

Most of published whole genome studies have been focused mainly on the coding part, which is only about 1.5 percent of the whole genome, because of knowledge that roughly 85% of the known genetic causes of Mendelian disorders affect the protein coding regions (8, 9). The term 'exome' can be defined as a coding part of genome – all exons. Whole exome sequencing (WES) is an effective strategy for selective sequencing of only coding regions of genome (9). Advantage using WES is because of two reasons: firstly, there are extremely rare diseases with only few affected individuals and families per disorder, which result in underpowered analyses and / or large regions under the linkage peak(s); secondly, despite that these disorders are rare, the causal mutations show the large phenotypic effect (2, 8).

Although many different methods for targeted capture have been described, only few have been extended to target the human exome. These highthroughput sequence capture methods are based on hybridization, for example, array-based hybridization or liquid-based hybridization depend on technology that is used (10).

DISCUSSION

Initial exome sequencing was made by Sarah Ng and colleagues in 2009, when gene of the rare autosomal recessive genetic disorder, Miller syndrome, was found (11). This new approach has opened a new revolutionary page of opportunities for the rare disease research and diagnostic. During brief 3 years of WES usage a considerably high number of novel recessive disease genes have been discovered. That includes all types of inheritable rare diseases - autosomal recessive, autosomal dominant, X-linked. WES allowed finding genes that have an autosomal dominant pattern in diseases such as Kabuki, Schinzel-Giedion, Hajdu-Cheney syndromes, paroxysmal kinesigenic dyskinesia, primary lymphoedema, high myopia, dilated cardiomyopathy, autism, and others (12–16).

New X-linked disease causing genes such as TARP syndrome, leucoencephalopathy or X-linked intellectual disability were also discovered (17–19). Using exome sequencing technique mutations were found in mitochondrial DNA genes such as *MRPL3* (mitochondrial cardiomyophathy) or *AARS2* (infantile mitochondrial cardiomyophathy) (13, 20). That shows a great impact of WES on rare disease diagnostics. The autosomal recessive diseases especially benefit from this new method, and this review concentrates on such diseases.

In diseases research, the best way to carry out research is by combining few methods. In case of

rare diseases, many authors combine homozigosity mapping and exome sequencing. In disease studies, exome sequencing is used both as a basis research and as a way to confirm that gene mutation actually causes a disease. Combined use of exome sequencing and homozygosity mapping was applied in the study of Van Den Ende-Gupta syndrome, Joubert syndrome and other disorders (21–23).

One of WES advantages is that disease genes may be identified using a very limited number of patients - they may be identified even from only one patient. For instance, polycystic kidney disease's genetical cause was identified from only one 5-year-old young boy who had this disease. With the help of exome sequencing, two heterozygous *PKHD1* gene mutations were found, which lead to substitution of an asparagine for an aspartate, and that is the cause of disease. In this study, the researchers combined both, family and population, strategies (24). Another example, a rare recessive *FLVCR2* gene mutation that causes Fowler syndrome and is associated with progressive destruction of central nervous system tissue, was found out by testing only two patients (25). Ability to identify disease causing genes by exome sequencing only from a single or small number of affected individuals has considerable importance, as some rare diseases are so rare that it is difficult to find enough patients for that disease research.

As it was mentioned, since November 2009, exome sequencing has led to significant progress of rare autosomal recessive Mendelian disease identification. In the Table, these diseases are divided into vision and hearing; skeletal, teeth and skin; ciliopathic; nervous system and neuromascular as well as immunological groups.

In a short period of time, WES made a huge impact on research of ciliopathies – an emerging class of human genetic disorders that is caused by defects in cilia. These defects in cilia are associated with a range of human diseases, such as primary ciliary dyskinesia, hydrocephalus, polycystic liver and kidney disease, and some forms of retinal degeneration (26). Using WES, mutations that cause ciliopathy diseases were found: Van Den Ende-Gupta, Sensenbrenner, Joubert, Bardet-Biedl syndromes, Leber congenital amaurosis, polycystic kidney disease, primary ciliary dyskinesia (21–24, 27–31). Exome sequencing effectively confirmed that the nonsynonymous mutation chr22:19115386 C>T in the SCARF2 gene, which is expressed during development, is responsible for Van Den Ende-Gupta syndrome that affects multi-system (21). Also there were studies where exome sequencing was applied to reveal genes causing Joubert syndrome - a neurological, ciliopathic disorder manifested by psychomotor retardation, hypotonia, and ataxia. In 2010, Edvardson (22) found mutation in the TMEM216 gene related to Joubert syndrome type 2 in Ashkenazi Jews, and in 2012 Srour (23) found other mutations in *C5orf42* gene that also cause Joubert syndrome in the French Canadian population. An autosomal-recessive disease that is characterized by sagittal craniosynostosis and facial, ectodermal, and skeletal anomalies is known as Sensenbrenner syndrome. Gilissen and colleges (27) sequenced exomes of 2 patients and indentified compound heterozygous mutations in exon 2 of the WDR35 gene, causing Sensenbrenner syndrome. Another disease's gene related with ciliopathy and affecting multi-system was identified in 2012. This disease is Bardet-Biedl syndrome and it is caused by a homozygous 5 bp deletion in the *LTZFL1* gene (28). Whole-exome sequencing performed by 2 different scientist teams showed that NMNAT1 gene mutation is causing ciliopathic disease Leber congenital amaurosis (29, 30). Polycystic kidney disease and primary ciliary dyskinesia are also ciliopathic diseases that were studied with the help of whole exome sequencing (24, 31).

Exome sequencing also has some impact on vision and hearing diseases studies. Stargardt macular dystrophy, Leber congenital amaurosis and retinitis pigmentosa are diseases that are related with vision disorders and all of them were successfully analyzed by whole exome sequencing (29–30, 32– 34). Stargardt macular dystrophy is an inheritable degeneration disease that causes progressive vision loss. Whole exome sequencing revealed 7 diseases likely causing variants across four genes, providing a confident genetic diagnosis in six previously uncharacterized participants. There were identified four previously missed mutations in the ABCA4 gene across three individuals. Also, mutations were identified in RDS/PRPH2, ELOVL, and CRB1 genes that are also likely to cause this disease (32). Retinitis pigmentosa (RP) is a heterogeneous group of

Disease	Primary affected systems	Location of mutation (gene)	Authors	Year
Vision and hearing diseases				
Stargardt macular dystrophy (STGD)	Vision	ABCA4	Storm et al. (32)	2012
Retinitis pigmentosa	Vision	DHDDS CYP4V2	Züchner et al. (33) Wang et al. (34)	2011 2012
Usher syndrome type 3	Vision, hearing	ABHD12	Eisenberger et al. (35)	2012
Nonsyndromic hearing loss	Hearing	GPSM2	Walsh et al. (39)	2010
Perrault syndrome	Hearing, genital	HSD17B4	Pierce et al. (36)	2010
Brown-Vialetto-van Laere syndrome	Hearing, muscle, central and	SLC52A3	Johnson et al. (37)	2010
	peripheral nervous system	(C20orf54)	Haack et al. (38)	2012
	Skeletal, teeth, skin diseas	ses		
Osteogenesis imperfecta	Skeletal	SERPINF1	Becker et al. (43)	2011
Skeletal dysplasia	Skeletal	POP1	Glazov et al. (41)	2011
3-M syndrome	Skeletal	CCDC8	Hanson et al. (42)	2011
Chondrodysplasia and abnormal joint development	Skeletal	IMPAD1	Vissers et al. (44)	2011
Miller syndrome	Skeletal, hearing	DHODH	Ng et al. (11)	2010
Progeroid syndrome	Skeletal, CNS*, hair, skin, eyes	BANF1	Puente et al. (57)	2011
Amelogenesis imperfecta	Teeth	FAM20A	O'Sullivan et al. (45)	2011
Kohlschütter-Tönz syndrome	Teeth, CNS	ROGDI	Schossig et al. (46)	2012
Peeling skin syndrome	Skin	CHST8	Cabral et al. (47)	2012
Kaposi's sarcoma	Skin	STIM1	Byun et al. (48)	2010
Ciliopathies				
Leber congenital amaurosis	Vision	NMNAT1	Perrault et al. (29) Chiang et al. (30)	2012 2012
Joubert syndrome type 2 (JBTS2)	CNS, skeletal,	THEM216	Edvardson et al. (22)	2010
Joubert syndrome (JBTS)	eyes, ears, kidney	C5orf42	Srour et al. (23)	2012
Bardet-Biedl syndrome	Skeletal, CNS, vision, kidney, liver, heart	LZTFL1	Marion et al. (28)	2012
Polycystic kidney disease	Kidney, liver, lungs, pancreas	PKHD1	Da et al. (24)	2012
Primary ciliary dyskinesia	Lings	numerous	$\frac{1}{\text{Berg et al}} (31)$	2011
Sensenbrenner syndrome	Skeletal, eyes, kidney, liver	WDR35	Gilissen et al. (27)	2011
Van Den Ende-Gunta syndrome	Skeletal CNS eves	SCARE2	Anastasio et al. (21)	2010
Van Den Ende Gapta syndrome	ervous system / Neuromascula	r diseases	7111d3td310 ct dl. (21)	2010
Nonsyndromic mental retardation	CNS	TECR	Caliskan et al. (49)	2011
Hyperphosphatasia mental retarda-	Grit	1201		2010
tion syndrome	CNS, skeletal, heart	PIGV PIGO	Krawitz et al. (50)	2012
Congenital cerebellar ataxia	Central and peripheral nervous system	GRM1	Guergueltcheva et al. (51)	2012
Infantile onset spinocerebellar ataxia	CNS, hearing	C10orf2	Dündar et al. (52)	2012
Spinocerebellar ataxia with psycho- motor retardation	Central and peripheral nervous system, muscle	SYT14	Doi et al. (53)	2011
Progressive external ophthalmoplegia	Central and peripheral nervous system, muscle, heart	RRM2B	Takata el al. (40)	2011
Lethal congenital contractural syn- drome Type 4 (LCCS4)	CNS, muscle, skeletal	MYBPC1	Markus et al. (54)	2012
Fowler syndrome	CNS	FLVCR2	Lalonde et al. (25)	2010
Immunological diseases				
Autoimmune lymphoproliferative syndrome	Blood	FADD	Bolze et al. (55)	2010
Aplastic anemia	Blood	MPL	Walne et al. (56)	2012

Table. Autosomal recessive disorders that were studied by using exome sequencing

 \star CNS – central nervous system.

progressive retinal degenerations. Its symptoms include night blindness, tunnel vision and bone-spicule pigmentation in retina. Recently, it was known that over 50 genes can cause RP, but that explains no more than half of the clinical cases. The rise of exome sequencing could give a new insight into retinitis pigmentosa. Good examples are Wang and his colleagues' research in a large Chinese family that allowed finding retinitis pigmentosa causative mutations in the *CYP4V2* gene (33), and Züchner and his team's work with the Ashkenazi Jewish family, which showed that the *DHDDS* gene also harbors retinitis pigmentosa causing mutation (34).

Hearing diseases were also studied by exome sequencing including Usher, Perrault and Brown-Vialetto-van Laere syndromes, nonsyndromic hearing loss (11, 35–39). Usher syndrome is a retinitis pigmentosa syndromic form when a patient not only has a condition of vision loss, but also has hearing loss. With the help of homozygosity mapping and next-generation targeted exons sequencing in the Lebanese family which has Usher syndrome type 3 phenotype, causative mutation was found in AB-HD12 gene (35). Perrault and Brown-Vialetto-van Laere syndromes are two more examples of hearing disorders investigated by exome sequencing. With the help of WES causative mutation in Perrault syndrome was identified in the HSD17B4 gene (36), and in Brown-Vialetto-van Laere syndrome such mutation was revealed in the SLC52A3 gene (37–38). Nonsyndromic hearing loss was also studied using WES (39).

WES also allowed finding disease genes whose products are functioning in mitochondria, but are encoded by nuclear genes. Such situation was determined for the *RRM2B* gene, associated with autosomal recessive progressive external ophthalmoplegia. The *RRM2B* gene encodes a small subunit of ribonucleotide reductase small 2-like protein p53R2 which plays the essential role in the maintenance of mtDNA (40).

With the aid of WES, many diseases that affect bones, teeth and skin were studied. That includes skeletal dysplasia, 3-M syndrome, osteogenesis imperfecta, chondrodysplasia and abnormal joint development, Miller syndrome – bone diseases; amelogenesis imperfecta, Kohlschütter-Tönz syndrome – teeth diseases; as well as skin diseases – peeling skin syndrome, Kaposi's sarcoma (11, 41–48). Skeletal dysplasia, that commonly is called dwarfism, is a group of disorders characterized by abnormalities of cartilage and bone growth. When whole-exome sequencing was applied to a family of two healthy parents and two affected children with skeletal dysplasia, two novel compound heterozygous loss-of-function mutations in the POP1 gene were found and identified as disorder causative mutations (41). Another investigation using WES related with skeletal dysplasia was held by Hanson et al. (42). Mutation in the CCDC8 gene was idenfitied as a cause of 3-M syndrome that is known as a syndromic form of skeletal dysplasia. One more genetic bone disorder is osteogenesis imperfecta. Patients with such disorder have very easily breaking bones. With the help of WES, it was determined that four unrelated individuals had SERPINF1 gene mutations as an osteogenesis imperfecta cause (43). As for teeth disease, amelogenesis imperfecta, characterized by abnormal enamel formation, and the related Kohlschütter-Tönz syndrome are examples of WES studied teeth diseases. Mutation in the FAM20A gene related with amelogenesis imperfecta (45) and the ROGDI gene mutation related with Kohlschütter-Tönz syndrome were found (46). One of successful examples for WES applying to the skin diseases is peeling skin syndrome, genetic disorder characterized by continual peeling of the skin. By homozygosity mapping and whole-exome sequencing a novel homozygous missense mutation was identified within the the CHST8 gene (47). Another example is finding that the gene STIM1 is related with Kaposi's sarcoma, which is skin cancer (48).

An important disorder group represents diseases that cause damage of the nervous system. In this category the whole-exome sequencing also keeps pace with other disorder groups and there is already not significant number of diseases inheritable in autosomal recessive or dominant manner and studied using WES. As for recessive diseases causing damage in the central nervous system and studied by WES, nonsyndromic mental retardation and hyperphosphatasia mental retardation syndrome are fine examples (49, 50). In these both diseases damage of CNS causes mental retardation. In 2011 using WES, Caliskan and colleagues found that mutation in the TECR gene causes nonsyndromic mental retardation (49). Krawitz using WES performed an exhaustive inquiry of hyperphosphatasia mental retardation syndrome and found mutations in two genes PIGV and PIGO causing this disease (50). Disorders related with ataxia were also studied with the help of WES. Ataxia develops as a consequence of cerebellum degeneration responsible for control of muscle coordination. With the aid of WES, different ataxia types were investigated – congenital cerebellar ataxia that is caused by mutation in the GRM1 gene, infantile onset spinocerebellar ataxia caused by mutation in the C10orf2 gene and spinocerebellar ataxia with psychomotor retardation caused by SYT14 gene's mutation (51–53). Previously mentioned progressive external ophthalmoplegia as well as lethal congenital contractural type 4 and Fowler syndromes also belong to the nervous system diseases group analyzed with WES (25, 40, 54).

Refering to autosomal recessive manner inheritable immunological diseases studied by whole-exome sequencing, autoimmune lymphoproliferative syndrome and aplastic anemia could be mentioned. Autoimmune lymphoproliferative syndrome is an immune system disorder characterised by too large number of lymphocytes production followed by numerous autoimmune problems. Previously it was known that FASL and FAS gene mutations cause this disorder, but with combination of whole-exome sequencing and genome-wide linkage analysis it was shown that lymphoproliferative syndrome also develops after mutation in the FADD gene (55). Another disease, aplastic anemia, when bone marrow fails to make enough blood cells, was studied using exome sequencing in the Tunisian family with 2 affected children. In both patients with aplastic anemia, the sequencing data showed the c.1248 G>A mutation in the MPL gene (56).

CONCLUSIONS

In conclusion, the whole-exome sequencing, during its short period of existing (three years), was successfully applied to many different rare autosomal recessive diseases. Such variety of diseases studied by WES are referred to in the present review. The number of studied diseases is further growing very fast; therefore WES will take an important place in rare disorders research. Further using of wholeexome sequencing and fast increasing information on the rare diseases allow prognosticating a wide application of WES in population studies.

ACKNOWLEDGEMENTS

This study is part of the LITGEN Project (VP1-3.1-MM-07-K-01-013) funded by the European Social Fund under the Global Grant Measure.

> Received 29 January 2013 Accepted 22 March 2013

References

- Dodge JA, Chigladze T, Donadieu J, Grossman Z, Ramos F, Serlicorni A, et al. The importance of rare diseases: from the gene to society. Arch Dis Child. 2011; 96: 791–2.
- 2. Bell CJ, Dinwiddie DL, Miller NA, Hateley SL, Ganusova EE, Mudge J, et al. Carrier testing for severe childhood recessive diseases by next-generation sequencing. Sci Transl Med. 2011; 3: 65ra4.
- Ott J, Kamatani Y, Lathrop M. Family-based designs for genome-wide association studies. Nat Rev Genet. 2011; 12: 465–74.
- Vissers LE, de Ligt J, Gilissen C, Janssen I, Steehouwer M, de Vries P, et al. A de novo paradigm for mental retardation. Nat Genet. 2010; 42: 1109–12.
- Carr IM, Diggle CP, Touqan N, Anwar R, Sheridan EG, Bonthron DT, et al. Identification of autosomal recessive disease loci using out-bred nuclear families. Hum Mutat. 2012; 33(2): 338–42.
- Wei X, Ju X, Yi X, Zhu Q, Qu N, Liu T, et al. Identification of sequence variants in genetic diseasecausing genes using targeted next-generation sequencing. PLoS ONE. 2011; 6(12): e29500.
- Majewski J, Rosenblatt DS. Exome and whole-genome sequencing for gene discovery: the future is now! Hum Mutat. 2012; 33(4): 591–2.
- Gilissen C, Hoischen A, Brunner HG, Veltman JA. Disease gene identification strategies for exome sequencing. Eur J Hum Genet. 2012; 20(5): 490–7.
- Majewski J, Schwartzentruber J, Lalonde E, Montpetit A, Jabado N. What can exome sequencing do for you? J Med Genet. 2011; 48(9): 580–9.
- Teer JK, Mullikin JC. Exome sequencing: the sweet spot before whole genomes. Hum Mol Genet. 2010; 19(R2): 145–51.
- 11. Ng SB, Buckingham KJ, Lee C, Bigham AW, Tabor HK, Dent KM, et al. Exome sequencing iden-

tifies the cause of a Mendelian disorder. Nat Genet. 2010; 42(1): 30–5.

- Ng SB, Nickerson DA, Bamshad MJ, Shendure J. Massively parallel sequencing and rare disease. Hum Mol Genet. 2010; 19(R2): 119–24.
- 13. Gilissen C, Hoischen A, Brunner HG, Veltman JA. Unlocking Mendelian disease using exome sequencing. Genome Biol. 2011; 12(9): 228.
- 14. Chen WJ, Lin Y, Xiong ZQ, Wei W, Ni W, Tan GH, et al. Exome sequencing identifies truncating mutations in *PRRT2* that cause paroxysmal kinesigenic dyskinesia. Nat Genet. 2011; 43(12): 1252–5.
- Shi Y, Li Y, Zhang D, Zhang H, Li Y, Lu F, et al. Exome sequencing identifies *ZNF644* mutations in High Myopia. PLoS Genet. 2011; 7(6): e1002084.
- 16. Ostergaard P, Simpson MA, Brice G, Mansour S, Connell FC, Onoufriadis A, et al. Rapid identification of mutations in *GJC2* in primary lymphoedema using whole exome sequencing combined with linkage analysis with delineation of the phenotype. J Med Genet. 2011; 48(4): 251–5.
- Johnston JJ, Teer JK, Cherukuri PF, Hansen NF, Loftus SK, Chong K, et al. Massively parallel sequencing of exons on the X chromosome identifies *RBM10* as the gene that causes a syndromic form of cleft palate. Am J Hum Genet. 2010; 86(5): 743–8.
- Tsurusaki Y, Osaka H, Hamanoue H, Shimbo H, Tsuji M, Doi H, et al. Rapid detection of a mutation causing X-linked leucoencephalopathy by exome sequencing. J Med Genet. 2011; 48(9): 606–9.
- Harakalova M, van den Boogaard MJ, Sinke R, van Lieshout S, van Tuil MC, Duran K, et al. Xexome sequencing identifies a *HDAC8* variant in a large pedigree with X-linked intellectual disability, truncal obesity, gynaecomastia, hypogonadism and unusual face. J Med Genet. 2012; 49(8): 539–43.
- Galmiche L, Serre V, Beinat M, Assouline Z, Lebre AS, Chretien D, et al. Exome sequencing identifies *MRPL3* mutation in mitochondrial cardiomyopathy. Hum Mutat. 2011; 32(11): 1225–31.
- Anastasio N, Ben-ORosenblatt DS, Majewski Jmran T, Teebi A, Ha KC, Lalonde E, et al. Mutations in SCARF2 are responsible for Van Den Ende-Gupta syndrome. Am J Hum Genet. 2010; 87(4): 553–9.
- 22. Edvardson S, Shaag A, Zenvirt S, Erlich Y, Hannon GJ, Shanske AL, et al. Joubert syndrome 2

(JBTS2) in Ashkenazi Jews is associated with a *TMEM216* mutation. Am J Hum Genet. 2010; 86(1): 93–7.

- 23. Srour M, Schwartzentruber J, Hamdan FF, Ospina LH, Patry L, Labuda D, et al. Mutations in *C5ORF42* cause Joubert syndrome in the French Canadian population. Am J Hum Genet. 2012; 90(4): 693–700.
- 24. Zhang D, Lu L, Yang HB, Li M, Sun H, Zeng ZP, et al. Exome sequencing identifies compound heterozygous *PKHD1* mutations as a cause of autosomal recessive polycystic kidney disease. Chin Med J. 2012; 125(14): 2482–6.
- 25. Lalonde E, Albrecht S, Ha KC, Jacob K, Bolduc N, Polychronakos C, et al. Unexpected allelic heterogeneity and spectrum of mutations in Fowler sindrome revealed by next-generation exome sequencing. Hum Mutat. 2010; 31: 918–23.
- Badano JL, Mitsuma N, Beales PL, Katsanis N. The ciliopathies: an emerging class of human genetic disorders. Annu Rev Genomics Hum Genet. 2006; 7: 125–48.
- 27. Gilissen C, Arts HH, Hoischen A, Spruijt L, Mans DA, Arts P, et al. Exome sequencing identifies WDR35 variants involved in Sensenbrenner syndrome. Am J Hum Genet. 2010; 87(3): 418–23.
- 28. Marion V, Stutzmann F, Gérard M, De Melo C, Schaefer E, Claussmann A, et al. Exome sequencing identifies mutations in *LZTFL1*, a BBSome and smoothened trafficking regulator, in a family with Bardet-Biedl syndrome with situs inversus and insertional polydactyly. J Med Genet. 2012; 49(5): 317–21.
- 29. Perrault I, Hanein S, Zanlonghi X, Serre V, Nicouleau M, Defoort-Delhemmes S, et al. Mutations in *NMNAT1* cause Leber congenital amaurosis with early-onset severe macular and optic atrophy. Nat Genet. 2012; 44(9): 975–7.
- 30. Chiang PW, Wang J, Chen Y, Fu Q, Zhong J, Chen Y, et al. Exome sequencing identifies *NMNAT1* mutations as a cause of Leber congenital amaurosis. Nat Genet. 2012; 44(9): 972–4.
- 31. Berg JS, Evans JP, Leigh MW, Omran H, Bizon C, Mane K, et al. Next generation massively parallel sequencing of targeted exomes to identify genetic mutations in primary ciliary dyskinesia: implications for application to clinical testing. Genet Med. 2011; 13(3): 218–29.
- 32. Strom SP, Gao YQ, Martinez A, Ortube C, Chen Z, Nelson SF, et al. Molecular diagnosis of putative

Stargardt disease probands by exome sequencing. BMC Med Genet. 2012; 13: 67.

- 33. Wang Y, Guo L, Cai SP, Dai M, Yang Q, Yu W, et al. Exome sequencing identifies compound heterozygous mutations in CYP4V2 in a pedigree with retinitis pigmentosa. PLoS One. 2012; 7(5): e33673.
- 34. Züchner S, Dallman J, Wen R, Beecham G, Naj A, Farooq A, et al. Whole-exome sequencing links a variant in *DHDDS* to retinitis pigmentosa. Am J Hum Genet. 2011; 88(2): 201–6.
- 35. Eisenberger T, Slim R, Mansour A, Nauck M, Nürnberg G, Nürnberg P, et al. Targeted nextgeneration sequencing identifies a homozygous nonsense mutation in *ABHD12*, the gene underlying PHARC, in a family clinically diagnosed with Usher syndrome type 3. Orphanet J Rare Dis. 2012; 7(1): 59.
- 36. Pierce SB, Walsh T, Chisholm KM, Lee MK, Thornton AM, Fiumara A, et al. Mutations in the DBP-deficiency protein *HSD17B4* cause ovarian dysgenesis, hearing loss, and ataxia of Perrault syndrome. Am J Hum Genet. 2010; 87(2): 282–8.
- Johnson JO, Gibbs JR, Van Maldergem L, Houlden H, Singleton AB. Exome sequencing in Brown-Vialetto-van Laere syndrome. Am J Hum Genet. 2010; 87(4): 567–9.
- Haack TB, Makowski C, Yao Y, Graf E, Hempel M, Wieland T, et al. Impaired riboflavin transport due to missense mutations in *SLC52A2* causes Brown-Vialetto-Van Laere syndrome. J Inherit Metab Dis. 2012; 35(6): 943–8.
- 39. Walsh T, Shahin H, Elkan-Miller T, Lee MK, Thornton AM, Roeb W, et al. Whole exome sequencing and homozygosity mapping identify mutation in the cell polarity protein *GPSM2* as the cause of nonsyndromic hearing loss DFNB82. Am J Hum Genet. 2010; 87(1): 90–4.
- 40. Takata A, Kato M, Nakamura M, Yoshikawa T, Kanba S, Sano A, et al. Exome sequencing identifies a novel missense variant in *RRM2B* associated with autosomal recessive progressive external ophthalmoplegia. Genome Biol. 2011; 12(9): R92.
- 41. Glazov EA, Zankl A, Donskoi M, Kenna TJ, Thomas GP, Clark GR, et al. Whole-exome re-sequencing in a family quartet identifies *POP1* mutations as the cause of a novel skeletal dysplasia. PLoS Genet. 2011; 7(3): e1002027.
- 42. Hanson D, Murray PG, O'Sullivan J, Urquhart J, Daly S, Bhaskar SS, et al. Exome sequencing iden-

tifies *CCDC8* mutations in 3-M syndrome, suggesting that *CCDC8* contributes in a pathway with *CUL7* and *OBSL1* to control human growth. Am J Hum Genet. 2011; 89(1): 148–53.

- 43. Becker J, Semler O, Gilissen C, Li Y, Bolz HJ, Giunta C, et al. Exome sequencing identifies truncating mutations in human *SERPINF1* in autosomal-recessive Osteogenesis imperfecta. Am J Hum Genet. 2011; 88(3): 362–71.
- 44. Vissers LE, Lausch E, Unger S, Campos-Xavier AB, Gilissen C, Rossi A, et al. Chondrodysplasia and abnormal joint development associated with mutations in *IMPAD1*, encoding the Golgi-resident nucleotide phosphatase, gPAPP. Am J Hum Genet. 2011; 88(5): 608–15.
- 45. O'Sullivan J, Bitu CC, Daly SB, Urquhart JE, Barron MJ, Bhaskar SS, et al. Whole-exome sequencing identifies *FAM20A* mutations as a cause of amelogenesis imperfecta and gingival hyperplasia syndrome. Am J Hum Genet. 2011; 88(5): 616–20.
- 46. Schossig A, Wolf NI, Fischer C, Fischer M, Stocker G, Pabinger S, et al. Mutations in *ROGDI* cause Kohlschütter-Tönz syndrome. Am J Hum Genet. 2012; 90(4): 701–7.
- 47. Cabral RM, Kurban M, Wajid M, Shimomura Y, Petukhova L, Christiano AM. Whole-exome sequencing in a single proband reveals a mutation in the *CHST8* gene in autosomal recessive peeling skin sindrome. Genomics. 2012; 99(4): 202–8.
- 48. Byun M, Abhyankar A, Lelarge V, Plancoulaine S, Palanduz A, Telhan L, et al. Whole-exome sequencing-based discovery of *STIM1* deficiency in a child with fatal classic Kaposi sarcoma. J Exp Med. 2010; 207(11): 2307–12.
- 49. Çalışkan M, Chong JX, Uricchio L, Anderson R, Chen P, Sougnez C, et al. Exome sequencing reveals a novel mutation for autosomal recessive non-syndromic mental retardation in the *TECR* gene on chromosome 19p13. Hum Mol Genet. 2011; 20(7): 1285–9.
- Krawitz PM, Schweiger MR, Rödelsperger C, Marcelis C, Kölsch U, Meisel C, et al. Identity-bydescent filtering of exome sequence data identifies *PIGV* mutations in hyperphosphatasia mental retardation syndrome. Nat Genet. 2010; 42(10): 827–9.
- 51. Guergueltcheva V, Azmanov DN, Angelicheva D, Smith KR, Chamova T, Florez L, et al. Autosomal-recessive congenital cerebellar ataxia is caused

by mutations in metabotropic glutamate receptor. Am J Hum Genet. 2012; 91(3): 553–64.

- 52. Dündar H, Ozgül RK, Yalnızoğlu D, Erdem S, Oğuz KK, Tuncel D, et al. Identification of a novel twinkle mutation in a family with infantile onset spinocerebellar ataxia by whole exome sequencing. Pediatr Neurol. 2012; 46(3): 172–7.
- 53. Doi H, Yoshida K, Yasuda T, Fukuda M, Fukuda Y, Morit H, et al. Exome sequencing reveals a homozygous *SYT14* mutation in adult-onset, autosomal-recessive spinocerebellar ataxia with psychomotor retardation. Am J Hum Genet. 2011; 89(2): 320–7.
- 54. Markus B, Narkis G, Landau D, Birk RZ, Cohen I, Birk OS. Autosomal recessive lethal congenital contractural syndrome type 4 (LCCS4) caused by a mutation in *MYBPC1*. Hum Mutat. 2012; 33(10): 1435–8.
- 55. Bolze A, Byun M, McDonald D, Morgan NV, Abhyankar A, Premkumar L, et al. Whole-exomesequencing-based discovery of human *FADD* deficiency. Am J Hum Genet. 2010; 87(6): 873–81.
- 56. Walne AJ, Dokal A, Plagnol V, Beswick R, Kirwan M, de la Fuente J, et al. Exome sequencing identifies *MPL* as a causative gene in familial aplastic anemia. Haematologica. 2012; 97(4): 524–8.

Tautvydas Rančelis, Loreta Cimbalistienė, Vaidutis Kučinskas

GENŲ, LEMIANČIŲ RETAS AUTOSOMINES RECESYVINES LIGAS, TYRIMŲ REZULTATAS PAGAL EGZOMO SEKOSKAITĄ

Santrauka

Reta laikoma ta liga, kuria serga tik nedidelė populiacijos dalis. Europos Sąjungoje reta liga laikoma tada, kai ja serga mažiau nei 1 iš 2 000 žmonių. Ir nors nuo vienos retos ligos kenčia nedaug žmonių, tokių retų ligų yra daug. Manoma, kad tokio pobūdžio ligų yra apie 7 000-8 000. Tad vien Europos Sąjungoje tokiomis ligomis sergančių asmenų priskaičiuojama per 30 milijonų. Toks didelis sergančių asmenų skaičius verčia domėtis ir tirti retas ligas, ir tai gali padėti išaiškinti šių ligų atsiradimo priežastis bei sukurti naujus, tobulesnius jų diagnozavimo būdus. Klasikinės paveldimų ligų tyrimo strategijos susiduria su papildomomis problemomis (dauginiai genetiniai variantai, de novo mutacijos, ypač reti atvejai), ir tai daro šias strategijas nepakankamai veiksmingas. Naujos kartos sekoskaita atveria naują retų ligų tyrimų puslapį. Viso egzomo sekoskaita per trumpą gyvavimo laikotarpį sugebėjo nemažai prisidėti ieškant retų autosominių recesyvinių ligų genetinių priežasčių – per trejus metus buvo aptikta ir ištirta daug naujų mutacijų genuose, lemiančių šias ligas.

Raktažodžiai: retos autosominės recesyvinės ligos, viso egzomo sekoskaita, sindromai